

## ALLYLPHENOLS FROM *OCOTEA CYMBARUM*\*

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**Key Word Index**—*Ocotea cymbarum*; Lauraceae; 4-hydroxy-2,3,5-trimethoxyallylbenzene; apiolglycol.

**Abstract**—An ethanol extract of *Ocotea cymbarum* wood was shown to contain apiol, dillapiol, 4-hydroxy-2,3,5-trimethoxyallylbenzene, apiolglycol and lyoni-resinol.

The wood of *Ocotea cymbarum* (H.B.K.) Nees has been shown to contain three allylphenol derivatives: eugenol, dehydrodieugenol, mono-*O*-methyldehydrodieugenol and dehydrodieugenol **B** [2]. In the present study the ethanolic wood extract of another specimen of this species collected in a similar locality near Manaus, Amazonas, was also found to contain allylphenol derivatives but of a much higher oxygenation pattern: apiol (**1a**), dillapiol (**1b**) [3], 4-hydroxy-2,3,5-trimethoxyallylbenzene (**1c**) and apiolglycol (**2**). Compound **1c**, the common putative precursor of **1a** and **1b**, is here described for the first time. Among the apiol, dillapiol, isoapiol and isodillapiol derived glycols only the latter has been isolated previously from *Ostericum citriodorum* (Apiaceae) [4]. Although a known synthetic derivative [5], **2** is thus a new natural product. The extract contained in addition the 4-aryltetralin type lignan lyoni-resinol, previously isolated from *Lyonia ovalifolia* (Ericaceae), *Alnus glutinosa* (Betulaceae) and *Ulmus thomasi* (Ulmaceae) [6].

Spectral comparison of **1a** and **1b** (both  $\text{ArH} \cdot \text{CH}_2\text{CH}=\text{CH}_2(\text{OMe})_2\text{O}_2\text{CH}_2$  by NMR and MS), **1c** [ $\text{ArH} \cdot \text{CH}_2\text{CH}=\text{CH}_2 \cdot \text{OH}(\text{OMe})_3$ ] and **2** [ $\text{ArH} \cdot \text{CH}_2\text{CH}(\text{OH})\text{CH}_2\text{OH}(\text{OMe})_2\text{O}_2\text{CH}_2$ ] led to the indicated structures via the following observations. The aromatic protons of **1c** and **2** can be *ortho*-related only with one oxy-group ( $^1\text{H}$ - $^{13}\text{C}$  NMR  $\delta$  **1a** 6.25/108.25; **1b**, 6.40/102.56; **1c** 6.42/107.11; **2** 6.30/109.27) as in **1a** and **1b**. As in **1a**, but not as in **1b**, a methoxyl must be vicinal to the sole free aromatic position in **1c** and in **2** ( $^{13}\text{C}$  NMR  $\delta$  **1a** 56.80; **1b** 61.08; **1c** 56.39; **2** 56.92). A minimal paramagnetic shift ( $\Delta 0.1$  ppm) of the ArH singlet occurs upon acetylation of the, hence, *meta*-related free hydroxyl of **1c**. Osmium tetroxide oxidation [7] of **1a** gives **2**.

### EXPERIMENTAL

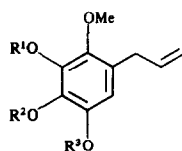
**Isolation of the constituents.** An EtOH extract of *Ocotea cymbarum* was kindly supplied by the Instituto Nacional de Pesquisas da Amazônia, Manaus, there registered as extract no. 407. Part of the extract (56 g) was re-extracted with EtOAc, the soln. evapd. and the residue (16 g) submitted to CC (silica gel). Elution with  $\text{CH}_2\text{Cl}_2$  gave, in order, a mixture of **1a**, **1b** and **1c** (purified by TLC silica gel,  $\text{CHCl}_3$ -MeOH 49:1), **1c** and sitosterol. Elution with  $\text{CH}_2\text{Cl}_2$ -MeOH 49:1 gave **2** (172 mg) (purified by recryst. from EtOAc). Another part of the extract (95 g) was submitted directly to CC (silica gel, 750 g). Elution with  $\text{CHCl}_3$  gave in order **1a** (30 g), **1b** (9 g), and **1c** (150 mg) (purified by TLC) and sitosterol (100 mg). Elution with  $\text{CHCl}_3$ -MeOH 19:1 gave lyoni-resinol (415 mg) (purified by recryst. from  $\text{Me}_2\text{CO}$ ).

**Lyoni-resinol**, a (8*R*,7'*S*,8'*R*)-8,8',6,7'-lignan (OH: 4,9,4',9'; OMe: 3,3',5,5';  $\Delta$ : 1,3,5,1',3',5') [8], mp 195–197° (Me<sub>2</sub>CO). Diacetate, mp 144–146°. Dimethyl ether, mp 168–170°.

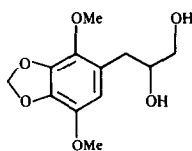
**4-Hydroxy-2,3,5-trimethoxyallylbenzene (1c).** Oil, UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm: 218 ( $\epsilon$  6900), 283 ( $\epsilon$  2050). IR  $\nu_{\text{max}}^{\text{film}}$   $\text{cm}^{-1}$ : 3450 (OH), 1650, 1600, 1500 (Ar), 990, 915 ( $\text{CH}=\text{CH}_2$ ).  $^1\text{H}$  NMR (60 MHz,  $\text{CDCl}_3$ )  $\delta$ : 3.31 (*d*,  $J$  = 7 Hz, 2H-7), 3.80, 3.86, 3.92, (3s, 3 OMe), 4.8–5.3 (2H-9), 5.6–6.2 (H-8), 5.57 (*s*, OH), 6.42 (*s*, H-6).  $^{13}\text{C}$  NMR (25 MHz,  $\text{CDCl}_3$ )  $\delta$ : 33.74 (*t*, C-7), 56.31 (*q*, OMe-5), 60.64, 60.98 (2*q*, 2OMe-2,3), 107.11 (*d*, C-6), 115.27 (*t*, C-9), 123.07 (*s*, C-1), 137.49 (*s*, C-4), 137.49 (*d*, C-8), 140.46 (*s*, C-3), 143.46 (*s*, C-2), 145.01 (*s*, C-5). MS  $m/z$  (rel. int.): 224 (*M*, 100), 209 (48), 195 (13), 117 (35), 163 (9), 149 (22), 121 (10). Acetate, oil, IR  $\nu_{\text{max}}^{\text{film}}$   $\text{cm}^{-1}$ : 1775 (OAc), 1655 ( $\text{CH}=\text{CH}_2$ ), 1615, 1490 (Ar), 984, 913 ( $\text{CH}=\text{CH}_2$ ).  $^1\text{H}$  NMR (60 MHz,  $\text{CDCl}_3$ )  $\delta$ : 2.28 (*s*, OAc), 3.42 (*d*,  $J$  = 7 Hz, 2 H-7), 3.80, 3.82, 3.90 (3*s*, 3 OMe), 4.9–5.3 (2 H-9), 5.7–6.1 (H-8), 6.55 (H-6).

**Apiolglycol (2).** Mp 100–101°. UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm: 216 ( $\epsilon$  6650), 280 ( $\epsilon$  550). IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3260 (OH), 1605, 1500, 1490 (Ar).  $^1\text{H}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 2.40 (*br s*, OH), 2.73 (*d*,  $J$  = 7 Hz, 2H-7), 3.4–3.6 (2 H-9), 3.84, 3.92 (2*s*, 2 OMe), 5.94 (*s*,  $\text{CH}_2\text{O}_2$ ), 6.30 (*s*, H-6).  $^{13}\text{C}$  NMR (25 MHz,  $\text{CDCl}_3$  +  $\text{C}_5\text{D}_5\text{N}$ )  $\delta$ : 34.46 (*t*, C-7), 56.92 (*q*, OMe-5), 59.81 (*q*, OMe-2), 65.92 (*t*, C-9), 72.41 (*d*, C-8), 101.27 (*t*,  $\text{CH}_2\text{O}_2$ ), 109.27 (*d*, C-6), 123.98 (*s*, C-1), 135.34 (*s*, C-4), 136.63 (*s*, C-3), 138.37 (*s*, C-2), 138.88 (*s*, C-5). MS  $m/z$  (rel. int.): 256 (*M*, 100), 238 (8), 225 (38), 196 (97), 195 (100), 181 (73), 180 (59), 165 (38), 151 (21), 137 (21), 135 (62), 109 (16). Diacetate, mp 110–112°.  $^1\text{H}$  NMR (60 MHz,  $\text{CDCl}_3$ )  $\delta$ : 2.08, 2.11 (2*s*, 2 OAc), 2.85 (*d*,  $J$  = 7 Hz, 2H-7), 3.90, 3.99 (2*s*, 2 OMe), 4.1–4.3 (2H-9), 5.2–5.5 (H-8), 6.01 (*s*,  $\text{CH}_2\text{O}_2$ ), 6.38 (*s*, H-6).

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- 1a**  $R^1-R^2 = CH_2$ ,  $R^3 = Me$   
**1b**  $R^1 = Me$ ,  $R^2 = R^3 = CH_2$   
**1c**  $R^1 = R^3 = Me$ ,  $R^2 = H$



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**Transformation of 1c in 2.** A soln of  $OsO_4$  (100 mg) in  $C_5H_5N$  (1 ml) was added to a soln. of **1c** (90 mg) in  $C_5H_5N$  (0.5 ml). After stirring (3 hr, room temp.) a soln of  $NaHSO_3$  (120 mg) in  $C_5H_5N$  (3 ml) and  $H_2O$  (2 ml) was added and the mixture stirred for 30 min before addition of 10% HCl (3 ml). Stirring continued for 30 min and the mixture then extracted with  $CHCl_3$ . The organic layer was washed, dried and evapd. The residue was purified (TLC, Si gel) to give **2**.

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## 4,4'-DIHYDROXYCHALCONE FROM THE HEARTWOOD OF *CHAMAECYPARIS OBTUSA*

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**Key Word Index**—*Chamaecyparis obtusa*; Cupressaceae; heartwood; phenolic extractives; 4,4'-dihydroxychalcone.

**Abstract**—A new chalcone, 4,4'-dihydroxychalcone was isolated from the heartwood of *Chamaecyparis obtusa*. The structure was elucidated by direct comparison with a synthetic sample.

## INTRODUCTION

Japanese cypress (*Chamaecyparis obtusa* Endl.), is highly valued for its pink heartwood. The phenolic extractives responsible for this colour are hinokinin, hinokiresinol, hinokione and hinokiol [1–7]. The present authors while reinvestigated the basis of this colour found, in addition to four known compounds (Sawaranin, cryptoresinol, 3-methoxyhinokiresinol and isocryptoresinol) [8–10], one new substance which is now described in this note.

## RESULTS AND DISCUSSION

The phenolic part of ethyl acetate-soluble fraction from the methanolic extract of the heartwood of *C. obtusa* was acetylated, and this eventually provided the acetate (**1b**) of the new compound, in a yield of 0.001% based on dried heartwood powder.

Upon preliminary TLC analysis of the original ethyl acetate fraction, compound **1a** appeared as a yellow spot which was positive to 2,4-dinitrophenylhydrazine and